

abia TG Ab



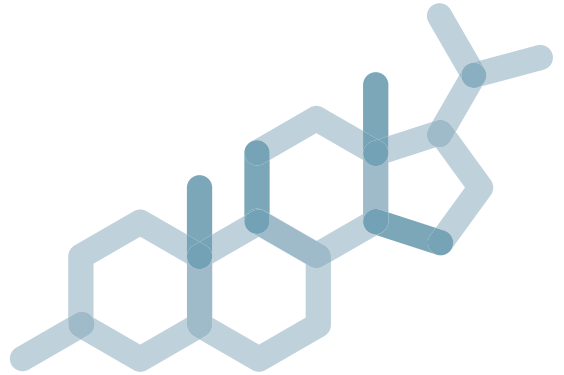
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Note: Changes highlighted ★

abia



Intended use

Abia TG Ab enzyme immunoassay for the quantitative determination of anti-thyroglobulin (anti-TG) autoantibodies in human serum.

The assay is intended for aid in the assessment of thyroid status and diagnosis of thyroid disease. For professional use only.

Clinical value

Thyroglobulin (TG), the principal storage protein normally present in the thyroid follicles, is composed of a 19S glycoprotein, thyroxine (T₄), triiodothyronine (T₃) and their precursors.

Thyroglobulin may spill into the circulation as a result of thyroid disease such as Hashimoto's thyroiditis, and cancer, and may deceive the immune system into producing anti-TG autoantibodies (anti-TG). Anti-TG autoantibodies belong mainly to the immunoglobulin G (IgG) class. The concentrations of circulating anti-TG autoantibodies vary over a wide range depending on the causative disease.

Therefore, quantitation of circulating anti-TG autoantibodies is important in the diagnosis as well as the follow-up of these thyroid diseases. Circulating anti-TG autoantibodies occur, in varying amounts, in patients suffering from such diseases as autoimmune thyroiditis due to Hashimoto's Disease, Graves' Disease, endemic goiter, subacute thyroiditis, and thyroid carcinoma. The differential diagnosis is further refined by measuring thyroid peroxidase autoantibodies, thyrotropin receptor autoantibodies (anti-TR autoantibodies) and thyroglobulin in serum.

Principle of the test

Abia TG Ab is a two-step indirect assay based on microwells coated with human thyroglobulin (TG). The conjugate is a mixture of HRP-labeled monoclonal anti-human-IgG antibodies.

Serum samples are added to the wells and if anti-TG antibodies are present in a sample, they form stable complexes with human TG immobilized on the wells.

Then the antigen-antibody complexes are identified by the addition of HRP labeled anti-human-IgG conjugate.

The unbound components are removed by washing. After addition of the solution containing TMB and hydrogen peroxide, the wells with bound conjugate develop a blue color which is converted to yellow after the reaction has been stopped with sulphuric acid.

The color intensity is directly proportional to the concentration of TG in the specimen and can be read at 450 nm.

Kit contents

	S	
TG Ag coated plate	1	polystyrene plate 12 × breakable 8-well strips coated with human thyroglobulin (TG)
Conjugate	1 × 12 ml	ready to use; HRP-labeled monoclonal anti-human IgG; transparent or slightly opalescent yellow liquid
Sample diluent 1	1 × 12 ml	buffer for the 1st samples dilution in preliminary plate; transparent or slightly opalescent blue-violet liquid
Sample diluent 2	1 × 12 ml	buffer for the 2nd samples dilution in TG Ag coated plate; transparent or slightly opalescent pink liquid
Calibrator 0	1 × 10 ml	protein based buffer not containing TG; pale yellow liquid
Calibrator 1	1 × 10 ml	human based buffer containing TG in concentration approx. 100 IU/ml; pale yellow liquid
Calibrator 2	1 × 10 ml	human based buffer containing TG in concentration approx. 250 IU/ml; pale yellow liquid
Calibrator 3	1 × 10 ml	human based buffer containing TG in concentration approx. 500 IU/ml; pale yellow liquid
Calibrator 4	1 × 10 ml	human based buffer containing TG in concentration approx. 1 000 IU/ml; pale yellow liquid
Calibrator 5	1 × 10 ml	human based buffer containing TG in concentration approx. 2 000 IU/ml; pale yellow liquid
Control serum	1 × 10 ml	human based buffer based control containing TG; pale yellow liquid
Washing solution (concentrated 25-fold)	1 × 50 ml	phosphate saline buffer; colorless or pale yellow liquid
TMB/substrate solution	1 × 12 ml	ready to use; citric acid buffer containing TMB and H ₂ O ₂ ; colorless liquid
Stopping reagent 0.2M H ₂ SO ₄	1 × 25 ml	ready to use; 0.20 mol/l sulphuric acid solution; colorless liquid
Protective film	2	
Plastic dish	2	
Plastic zip-lock bag	1	

The calibrators were calibrated using a 1st International Standard 65/093. Exact concentration levels for calibrators and control serum are given on the labels and certificates of analysis on a lot specific basis. All components are stable until expiration date of the kit when stored at 2–8 °C in a tightly sealed package. Expiration date is indicated on the package. Once opened, the components should be used within one month. Concentration of preserving agents: <=0.1 %.

Materials and equipment required but not provided

- purified water
- automatic or semiautomatic, adjustable or preset pipettes or multipipettes
- disposable pipette tips
- preliminary plate for samples pre-dilution with the sample diluent 1 reagent
- automatic microplate washer
- microplate reader equipped with 450 and 405-415 nm filter

Safety notes

- human origin material used in the preparation of the calibrators and control serum has been tested by CE-marked tests and found non reactive for hepatitis B surface antigen (HBsAg), antigen p24 HIV-1, antibodies to hepatitis C virus and antibodies to human immunodeficiency virus (HIV-1 and HIV-2)
- as no known test method can offer complete assurance that infectious agents are absent, reagents and samples should be handled as if capable of transmitting infectious disease; any equipment directly in contact with samples and reagents should be considered as contaminated
- do not eat, drink, smoke or apply cosmetics in the laboratory
- do not pipette by mouth
- avoid any contact of the reagents and samples with the skin and mucosa; wear lab coats and disposable gloves when handling them; thoroughly wash your hands after work
- avoid spilling samples or solutions containing samples. Wipe spills immediately and decontaminate affected surfaces
- all materials contacted with specimens or reagents, including liquid and solid waste, should be inactivated by validated procedures (autoclaving or chemical treatment) and disposed in accordance with applicable local law regulations

Precautions

- do not use reagents without label or with damaged label/package
- do not use expired reagents
- do not change the assay procedure; perform all subsequent steps without interruption
- do not mix reagents from different lots
- do not mix the caps of vials
- do not run the EIA test in the presence of reactive vapours (acid, alkaline, aldehyde), dust or metals
- do not let the wells dry once the assay has been started
- do not use the same container and tips for different liquid components of the kit and samples
- do not reuse the coated plates
- do not reuse the removed protective film
- do not expose the reagents to excessive heat or sunlight during storage and test procedure
- do not freeze the reagents

Collection and handling of specimens

- collect blood specimens according to the current practices
- use serum for testing; performances of the test have not been evaluated on other biological fluids
- separate the clot or red cells from serum as soon as possible to avoid any haemolysis
- do not use sera preserved with sodium azide, thiomersal, phenol
- do not use contaminated, hyperlipaemic and hyperhaemolysed specimens
- the samples with hyperproteinaemia and hyperbilirubinaemia were not specially tested
- before testing samples with observable particulate matter should be clarified by centrifugation
- suspended fibrin particles or aggregates may yield reactive results
- do not heat the samples
- samples can be stored at 2–8 °C within 72 hours or deep-frozen at -20 °C
- no more than one freeze/thaw cycle is allowed

Procedural notes

- before use wait 30 minutes for the reagents to stabilize to room temperature (20–25 °C)
- check appearance of the reagents
- lost vacuum in the bag of the coated plate will not affect the performance of the test
- check the pipettes and other equipment for accuracy and correct operation
- the washing procedure is a critical step; for the detailed washer settings see section “Washing procedure”
- for the description of test procedure with the automated analyzers see section “Automated analyzers”

Washing procedure

Please contact your representative for protocols for recommended washers and procedures. In general the following protocol is recommended:

- flow-through washing with a volume not less than 300 µl per well is used
- repeat 3 times in the step 5 and 5 times in the step 8 of the test procedure
- do not allow the wells to become dry during the assay procedure
- ensure that no liquid is left in the well (use double aspiration in the final step where possible)
- avoid to tap out the plate
- residual volume lower than 10 µl is not critical for following steps of the test procedure
- when using a microplate washer clean the wash head frequently to prevent contamination

Preparation of reagents

Number of strips to be used	1	2	3	4	5	6	7	8	9	10	11	12
Working washing solution: mix the reagents thoroughly by inversion Stability: 14 days at 18–24 °C or 28 days at 2–8 °C												
Washing solution (concentrated 25-fold), ml	3.0	6.0	9.0	12.0	15.0	18.0	21.0	24.0	27.0	30.0	33.0	40.0
Purified water, ml	72.0	144.0	216.0	288.0	360.0	432.0	504.0	576.0	648.0	720.0	792.0	960.0

Test procedure

abia TG Ab for the quantitative determination of anti-thyroglobulin (anti-TG) autoantibodies concentration in human serum

- 1 Take the required number of coated strips. Place the unused strips back into the bag; reseal the foil-lined package in zip-lock plastic bag. Do not remove desiccant.
 - 2 Add 90 μ l of sample diluent 1 to preliminary plate (not provided).
Add 10 μ l of samples to be tested into appropriate wells.
Mix the contents of the wells by gentle pipetting. Blue-violet color should change to blue-green. No change of the color can be observed if no serum added to the well.
 - 3 Analyse each calibrator, control serum and samples in duplicate.
Add 100 μ l of calibrators 0 - 5, control serum into appropriate wells.
Add 90 μ l of sample diluent 2 in rest of the wells.
Add 10 μ l of prediluted samples to be tested from preliminary plate in the wells with sample diluent 2. Final dilution ratio is 1:100.
Mix the contents of the wells by gentle pipetting, then cover the plate with protective film.
The total time should not exceed 10 min.
 - 4 Incubate for 60 minutes at room temperature 20–25 °C.
 - 5 Remove the protective film slowly and carefully to prevent splashes. Aspirate the contents of all wells into a container for biohazardous waste (containing disinfectant).
Add not less than 300 μ l of working washing solution into each well and aspirate. Perform this procedure 3 times. Use double aspiration in the final step where possible.
 - 6 Add 100 μ l conjugate into each well.
Mix the contents of the wells for 30 seconds by careful tapping on the edge of the plate, then cover the plate with protective film.
 - 7 Incubate for 60 minutes at room temperature 20–25 °C.
 - 8 Remove the protective film slowly and carefully to prevent splashes. Aspirate the contents of all wells into a container for biohazardous waste (containing disinfectant).
Add not less than 300 μ l of working washing solution into each well and aspirate. Perform this procedure 5 times. Use double aspiration in the final step where possible.
 - 9 Add 100 μ l of TMB/substrate solution to all the wells. Keep the plates in a dark place for 10–20 minutes at 20–25 °C.
 - 10 Add 150 μ l of stopping reagent into each well. Mix gently for 5–10 sec.
 - 11 Read the optical density at 450 nm using a plate reader within 20 minutes after stopping reaction.
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Automated analyzers

Validated protocols for automated analyzers can be obtained from your representative. For the instrumentation without established validated protocol follow section “Test procedure” and ensure all requirements described in section “Precautions” are followed. All protocols for automated analyzers must be fully validated prior usage.

Calculation and interpretation of the results

Assay validation

Results of an assay are valid if the following criteria for the controls are met.

The absorbance (OD) of calibrator 5 should be greater than 1.300.
The absorbance (OD) of control serum should be within established range.

Calculation procedure

- 1 Calculate the mean optical density of each calibrator duplicate.
- 2 Calculate the mean optical density of each samples duplicate.
- 3 Draw a calibration curve on linear graph paper with the mean optical densities on the Y-axis and the calibrator concentrations on the X-axis.
- 4 Read the values of the unknowns directly off the calibration curve.
If immunoassay software is being used a 4-parameter curve is recommended.

Example	OD 1	OD 2	Mean OD	Value, IU/ml
Calibrator 0	0.040	0.036	0.038	0
Calibrator 1	0.301	0.299	0.300	125
Calibrator 2	0.540	0.505	0.522	300
Calibrator 3	1.053	1.024	1.036	550
Calibrator 4	2.100	1.915	1.965	1 100
Calibrator 5	2.834	2.816	2.824	2 100
Sample	0.915	0.974	0.945	506

This data is for illustration only and should **not be used** to calculate of samples. Each user should obtain his or her own data and standard curve.

Performance characteristics

Analytical sensitivity

The analytical sensitivity (limit of detection) was calculated by determining the variability of the calibrator 0 based on 12 analysis runs additional 2 x SD. Limit of detection defined at 10 IU/ml.

Specificity

No cross-reactivity to ANA, DNA, thyroid peroxidase (TPO) and rheumatoid antibodies was observed with this assay.

Precision	Mean, IU/ml	SD	CV, %
Intra-assay, sample 1	382	12.13	3.20
Inter-assay, sample 1	379	21.92	5.80

Accuracy

The assay was compared with a other immunoassay as a reference test. The total number of specimens was 162. The values ranged from 0 to 2872 IU/ml. The correlation coefficient were computed for abia TG Ab in comparison with the reference method. The correlation coefficient is 0.988.

Expected normal value

A normal range of less than 100 IU/ml anti-TG was obtained by testing serum samples from 250 individuals determined as normal by abia TSH and abia fT4 assays.

Normal value ranges may vary slightly among different laboratories. It is strongly recommended that each laboratory should determine its own range of expected normal values.













Limitations of test

- assay was validated only for the determination of anti-TG antibodies in human serum
- the results obtained with this assay should never be used as the sole basis for clinical diagnosis. Any laboratory result is only a part of the total clinical picture of the patient
- the presence of autoantibodies to TG is confirmed when the serum level exceeds 100 IU/ml. The clinical significance of the result, coupled with anti-thyroglobulin activity, should be used in evaluating the thyroid condition. However, clinical inferences should not be solely based on this test but rather as an adjunct to the clinical manifestations of the patient and other relevant tests

References

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2. Vole R, Clin Chem, Vol. 40, 2132 (1994).
3. Anderson, J.W, et al.: "Diagnostic Value of Thyroid Antibodies" J Clin Endocrinol Metab; 37:937-944 (1987).
4. Burek, C.L. and Rose, N.R.: "Detection of autoantibodies" in Gradwaohl's Clin Lab Methods and Diagnosis, Ed.8, C.V. Mosby, St. Louis, MO, pp. 1275-1278, (1978).

Key to symbols used

	Manufacturer
	For in vitro diagnostic use
	Catalogue number
	Batch code
 YYYY-MM-DD	Expiry date
 2°C - 8°C	Storage temperature limitation
	Do not use if package is damaged
	Do not reuse
 Σ_n	Sufficient for [n] tests
	Consult Instructions for use
	Caution, consult documents
	Changes highlighted

Hazard and precautionary statements for certain kit components

Stopping reagent



Warning

H315	Causes skin irritation.
H319	Causes serious eye irritation.
P264	Wash hands thoroughly after handling.
P280	Wear protective gloves/protective clothing/eye protection/face protection.
P302 + P352	IF ON SKIN: Wash with plenty of soap and water.
P305 + P351 + P338	IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

Conjugate



Warning

H317	May cause an allergic skin reaction.
P261	Avoid breathing dust/fume/gas/mist/vapours/spray.
P280	Wear protective gloves/protective clothing/eye protection/face protection.
P302 + P352	IF ON SKIN: Wash with plenty of soap and water.
P333 + P313	If skin irritation or rash occurs: Get medical advice/attention.

Attention!

For complete precautionary statements and detailed information see safety data sheets (SDS).



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