

**REF** DK.060.01.8  96

**IVD** For *In vitro* Diagnostic Use

**INSTRUCTIONS FOR USE**  
**abia CMV IgM**  
**Enzyme immunoassay for the detection**  
**of IgM antibodies to**  
***Cytomegalovirus* (CMV) in human serum or plasma**

**This Package Insert provides information for Professional Use of the kit.**

The kit contains sufficient reagents for 96 (one breakable plate) assays including controls; the kit is intended for manual testing with a possibility of fractional (one strip) use of the kit or for use of the kit on open type automated analyzer for enzyme immunoassay.

## **I. INTENDED USE**

The abia CMV IgM kit is intended for the detection of IgM antibodies specific to *Cytomegalovirus* (CMV) in human serum (plasma) by a microplate immunoenzymometric assay.

This kit is for diagnostic use by a trained laboratory professional and will not be sold to the general public. All the reagents are for professional in vitro diagnostic use only.

**The results of this or any other diagnostic assay should be used and interpreted only in the context of the overall clinical picture.**

## **II. INTRODUCTION**

*Cytomegalovirus* (CMV) is a member of the human herpesvirus group. Transmission occurs by contact to body fluids (saliva, genital secretions, urine, breast milk) and vertically in utero or during delivery. CMV can also be transmitted by blood transfusion, transplantation of organs and stem cells [1].

The presence of anti-CMV IgM may indicate one of the following: primary infection, re-infection, reactivation. The highest risk of infection of the fetus is observed in the primary infection in the mother during pregnancy (30-40%) [2]. Of all pregnancies with confirmed vertical transmission, only 10% to 20% of the fetuses will have evidence of clinical infection at birth [3]. For the diagnosis of primary CMV infection during pregnancy the detection of CMV antibody IgM with low IgG levels is shown.

For CMV infection is characteristic: 1) IgM may persist for many months after the primary CMV infection; 2) IgM can be detected during a secondary infection; 3) there may be cross reactivity with IgM due to another viral infection, for example, Epstein-Barr virus; 4) IgM can be detected as a result of nonspecific polyclonal stimulation of the immune system [4].

## **III. PRINCIPLE OF THE TEST**

Scheme of the test procedure is an indirect two-stage immunoassay. Microtiter strip wells precoated with the recombinant antigens of CMV to bind corresponding antibodies. The antigen-antibody complex reacted with HRP-labeled anti-human-IgM antibodies. The presence of bound enzyme indicating the presence in the specimen of specific antibodies is revealed by a color change in the TMB-Substrate. The color development is stopped with the addition of Stopping Reagent, changing the color to yellow. The color intensity of the test sample is directly proportional to the concentration of anti-CMV IgM.

## IV. CONTENT OF THE KIT abia CMV IgM

### 4.1 Contents of the reagent kit.

Table 1

LABEL	NATURE OF THE REAGENTS	PRESENTATION
CMV-Ag Coated Strips	Polystyrene stripped 96-well plate (breakable wells) coated with a mix of recombinant proteins, which represent the recombinant analogs of CMV antigens. Store at 2-8 °C until expiration date.	1 plate
Conjugate	Anti-human IgM polyclonal antibodies, conjugated with HRP enzyme with addition of urea (16.74%), 1M Tris HCl buffer (pH 7.35-7.45), Tween®20 (0.10%). Preserving agents: 0.10% ProClin 300, 0.004% gentamicin sulfate. Transparent or slightly opalescent yellow colored liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 11.0 ml
Positive Control, Inactivated	Control sample, containing Anti-GST antibodies, conjugated with HRP enzyme with addition of bovine serum albumin (1.95%), 1M Tris HCl buffer (pH 7.4-7.6), Tween®20 (0.11%). Preserving agents: 0.10% ProClin 300, 0.01% phenol. Transparent or slightly opalescent red colored liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 1.2 ml
Negative Control, Inactivated	Control sample, not containing IgM antibodies to CMV. Preserving agents: 0.04% ProClin 300, 0.2% sodium azide, 0.001% gentamicin sulfate. Transparent or slightly opalescent green colored liquid. Store at 2-8 °C until expiration date in a tightly sealed vial	1 vial 2.5 ml
Preliminary Sample Diluent	Sample buffer that is used for preliminary dilution of samples. Preserving agent: 0.09% sodium azide. Transparent or slightly opalescent violet-blue colored liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 11.0 ml
Sample Diluent	Sample buffer that is used to dilute samples before analysis with addition of bovine serum albumin (1.97%) and Tween®20 (0.11%). Preserving agent: 0.10% ProClin 300. Transparent or slightly opalescent pink colored liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 11.0 ml
Washing Solution (concentrated 25-fold)	Phosphate-saline solution (pH 7.4-7.7). Transparent or slightly opalescent colorless or light yellow liquid, sediment may form that dissolves completely at 35-39 °C and shaking. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 50.0 ml
Stopping Reagent	Sulfuric acid solution (H <sub>2</sub> SO <sub>4</sub> ) 0.2M. Transparent colorless liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 25.0 ml
TMB-Substrate	Tetramethylbenzidine in citric acid buffer, containing H <sub>2</sub> O <sub>2</sub> . Transparent colorless liquid. Store at 2-8 °C until expiration date in a tightly sealed vial.	1 vial 14.0 ml
Plate for preliminary dilution of sera	Polystyrene plate with transparent wells.	1 plate
Protective films for EIA plates		2
Polyethylene bag with a Zip-Lock		1
Disposable plastic dishes for liquid reagents		2
Disposable tips		16

## V. PRECAUTIONS

The reliability of the results depends on correct implementation of the following requirements:

- The temperature in the lab should be 18-24 °C.
- Inspect the contents of the box: check the vials and labels integrity. If labels are lost or labels/vials damage, vials should be disposed, and **kit cannot be used**.
- Do not use expired reagents.
- Do not mix reagents from different lots within a given test run.
- Do not carry out the test in the presence of reactive vapors (acid, alkaline, aldehyde vapors) or dust that could alter the enzyme activity of the conjugates.
- Use glassware thoroughly washed and rinsed with deionized water or preferably, disposable material.
- Do not allow the microplate to dry between the end of the washing operation and the reagent distribution.
- The enzyme reaction is very sensitive to metal ions. Consequently, do not allow any metal element to come into contact with the various conjugate or substrate solutions.
- Use a new distribution tip for each sample.
- Well washing is a critical step in this procedure: respect the recommended number of washing cycles and make sure that all wells are completely filled and then completely emptied. Incorrect washing may lead to inaccurate results.
- Never use the same container to distribute conjugate and color development solution.
- Check the pipettes and other equipment for accuracy and correct operation.
- Do not change the assay's procedure.
- Use high quality water.
- Avoid exposure of the reagents to excessive heat or sunlight during storage and incubation.

## VI. HEALTH AND SAFETY INSTRUCTIONS

- All reagents included in the kit are intended for “in vitro diagnostic use”.
- Human origin material used in the preparation of Negative Control has been tested and found negative for HBsAg, antibodies to hepatitis C virus and antibodies to human immunodeficiency virus (HIV-1 and HIV-2), antigen p24 HIV-1.
- Because no known test method can offer complete assurance that infectious agents are absent, handle reagents and patients samples as if capable of transmitting infectious disease.
- Do not eat, drink, smoke, or apply cosmetics where immunodiagnostic materials are being handled.
- Any equipment directly in contact with specimens and reagents as well as washing solutions should be considered as contaminated products and treated as such.

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- Wear lab coats and disposable gloves when handling reagents and samples and thoroughly wash your hands after handling them.
- Avoid spilling samples or solutions containing samples. Wipe spills immediately and decontaminate affected surfaces.
- Avoid any contact of the TMB-Substrate and the Stopping Reagent with the skin and mucosa.
- Provide adequate ventilation.
- All materials contacted with specimens or reagents, including liquid and solid wastes, should be inactivated by validated procedures (autoclaving or chemical treatment) and disposed in accordance with applicable local law regulations.



**Warning!**

Conjugate, Positive Control and Sample Diluent contain ProClin 300.

H317: May cause an allergic skin reaction.

P261: Avoid breathing vapours.

P280: Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352: IF ON SKIN: Wash with plenty of water.

P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.



**Danger!**

Negative Control contains sodium azide.

H312: Harmful in contact with skin.

P280: Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 IF ON SKIN: Wash with plenty of soap and water.

P312: Call a POISON CENTER or doctor/physician if you feel unwell.



**Danger!**

Stopping Reagent contains 0.2 M/l sulfuric acid.

H314 Causes severe skin burns and eye damage.

P280 Wear protective gloves/protective clothing/ eye protection/face protection.

P303 + P361 + P533 IF ON SKIN (or hair): Remove/Take off immediately all contaminated clothing. Rinse skin with water/shower.

P305 + P351 + P338 IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P310 Immediately call a POISON CENTER or doctor/ physician.

## **VII. MATERIALS AND EQUIPMENT REQUIRED BUT NOT PROVIDED WITH THE KIT:**

- Distilled or deionized water.
- Automatic or semiautomatic, adjustable or preset single-channel and multi-channel pipettes with a changeable volume for a set of liquids.
- Disposable pipette tips.
- Microplate incubator at  $(37.0 \pm 1.0) ^\circ\text{C}$ .

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- Automatic microplate washer.
- Microplate reader equipped with 450 nm and 620-680 nm filters.
- Laboratory clock.
- Open type automated analyzer with 450 nm and 620-680 nm filters (for automated procedure).

## **VIII. COLLECTION AND HANDLING OF SPECIMENS**

Collection of blood samples should be implemented according to the current practices. Serum, plasma (citrate, heparin, EDTA) may be used. Separate serum or plasma from blood cells as soon as possible to avoid any haemolysis. Extensive haemolysis may affect test performance. Specimens with observable particulate matter should be clarified by centrifugation prior to testing. Suspended fibrin particles or aggregates may yield falsely positive results. The samples after heat inactivation cannot be analyzed.

Samples can be stored at 2-8 °C not more than for 48 hours; they may be deep-frozen at -20 °C. Samples that have been frozen and defrosted more than 1 time cannot be used.

## **IX. PREPARATION OF THE REAGENTS**

### **1. Ready to use reagents:**

- **CMV-Ag Coated Strips.** Strips are wrapped in a sealed foil-lined bag. Open the bag and remove the tray. Select the number of Coated Strips required for the assay. Return unused strips in the bag. After the bag has been opened the Coated Strips are stable during the shelf life of the kit at 2-8 °C, provided that the foil-lined bag is resealed in Zip-Locked plastic bag. The silica gel bag should not be removed from the foil packaging.
- **Conjugate;**
- **Positive Control;**
- **Negative Control;**
- **Preliminary Sample Diluent;**
- **Sample Diluent;**
- **Stopping Reagent;**
- **TMB-Substrate.**

### **2. Reagents to prepare:**

- **Working Washing Solution.** Thoroughly shake Washing Solution concentrate. To make Working Washing Solution take required amount of concentrate and mix with distilled or deionized water (1:24 ratio) in a separate vial. Thoroughly mix the solution. The prepared Working Washing Solution is stable for 14 days at 18-24 °C or for 28 days at 2-8 °C.

The required volumes of Working Washing Solution for the certain number of strips or plate are tabulated in Table 2.

Table 2

Number of strips to be used		1	2	3	4	5	6	7	8	9	10	11	12	1 well
<b>Working Washing Solution</b>	Washing Solution (×25), ml	3.0	6.0	9.0	12.0	15.0	18.0	21.0	24.0	27.0	30.0	33.0	40.0	0.2
	High quality water, ml	72.0	144.0	216.0	288.0	360.0	432.0	504.0	576.0	648.0	720.0	792.0	960.0	4.8

### 3. Storage of unused reagents

After opening the vials the unused components of the kit: Positive Control, Negative Control, Preliminary Sample Diluent, Sample Diluent, Washing Solution (concentrated 25-fold), Stopping Reagent can be stored in tightly sealed vials until the kit expiration date at 2-8 °C. Coated Strips can be stored until the kit expiration date at 2-8 °C.

Conjugate, TMB-Substrate can be stored in tightly sealed vials within 3 months at 2-8 °C.

## X. TEST PROCEDURE

**Note: Before use, allow reagents to reach room temperature (18-24 °C) for 30 min.**

Step	The assay procedure
1	Add 90 µl of Preliminary Sample Diluent into the wells of the plate for preliminary samples dilution and 10 µl of the serum (plasma) samples. Carefully mix by pipetting. Violet-blue color should change to blue-green.
2	Add 100 µl of Positive and Negative Controls into the wells. <u>1 strip</u> – Positive Control to 1 well and Negative Control to 2 wells; <u>2 strips</u> and more – Positive Control to 1 well and Negative Control to 3 wells.
3	Add 90 µl of Sample Diluent and 10 µl of the preliminary diluted samples to the rest of the wells (the final serum dilution ratio is 1:100). Carefully mix fluid in wells by gentle pipetting. Cover the strips with a protective film.
4	Incubate for <b>30 min</b> in a microplate incubator at (37.0 ± 1.0) °C.
5	Aspirate the contents of the wells and wash the plate <b>4 times</b> with the Working Washing Solution. Add into each well not less than 380 µl of Working Washing Solution and remove Washing Solution into the container with disinfecting solution. Do not leave any fluid in the wells. Use of an automatic microplate washer is strongly recommended. Incomplete washing will adversely affect the assay precision.
6	Add 100 µl of Conjugate to all the wells of the plate. Cover the plate with a protective film.
7	Incubate for <b>30 min</b> in a microplate incubator at (37.0 ± 1.0) °C.
8	Remove fluid from wells, wash the plate <b>4 times</b> as described in step 5.
9	Add 100 µl of TMB-Substrate into all the wells.
10	Incubate at 18-24 °C for a <b>20 min</b> in a dark place.

11	Add 150 µl of Stopping Reagent into wells to stop the reaction results are read by microplate plate reader at wavelength of 450 nm, with reference filter at 620-680 nm. In case of overflow absorbance values, read at 405 nm, with reference filter at 620-680 nm.
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Scheme of the assay is represented in Annex.

### Automated analyzer

Validated test protocols and dilution tables of reagent working solutions for different EIA-analyzers can be obtained from the manufacturer upon request (see section XIV). For the instrumentation without established validated protocol follow the section “TEST PROCEDURE” and ensure all requirements described in the section “PRECAUTIONS” are fulfilled. All protocols for automated analyzers must be fully validated before use.

When preparing working reagent solutions for automated EIA procedure, it is necessary to consider “dead” volume of vials and containers used for loading working solutions in the EIA analyzer.

## XI. RESULTS

### 1. Test Validation

For the test to be valid the following criteria must be met. If these criteria are not met the test should be considered invalid and should be repeated.

1. **Positive Control:** the absorbance value should not be less than 1.0;
2. **Negative Control:** the absorbance value should not be more than 0.2.

Calculate Cut-Off value as:

$$\text{Cut-Off} = \text{average OD value of Negative Control} + A, \quad (A=0.150)$$

where **A** – is a coefficient defined by manufacturer during statistical processing for each lot.

### 2. Interpretation of Results

Sample is positive, if the OD value is  $\geq$  Cut-Off.

Sample is negative, if the OD value is  $<$  Cut-Off.

## XII. PERFORMANCE CHARACTERISTICS

### 1. Interferences

Hemoglobin (up to 28.32 mg/ml), bilirubin (up to 0.3 mg/ml), lipids (up to 11.4 mg/ml), rheumatoid factor (up to 221 IU/ml) have no influence on the assay results.

### 2. Cross reactivity

No cross reactivity was found for samples with antibody to *Rubella virus*, *Epstein-Barr virus* (EBV).



### 3. Diagnostic sensitivity

Diagnostic sensitivity of abia CMV IgM with 60 anti-CMV IgM positive samples is 93.3% (95%CI: 84.1-97.4).

### 4. Diagnostic specificity

Diagnostic specificity of abia CMV IgM with 1527 anti-CMV IgM negative samples is 97.3 % (95% CI: 96.3-98.0).

### 5. Trueness. Agreement with certified reference measurement procedure

The abia CMV IgM was compared with the “SERION ELISA classic Cytomegalovirus IgM”, Virion/Serion. 153 serum and plasma samples are tested.

abia CMV IgM	“SERION ELISA classic Cytomegalovirus IgM”			
		Positive	Negative	Indeterminate
	Positive	74	18	6
Negative	15	37	3	

The abia CMV IgM has not a “gray area”, so indeterminate results were not included in the calculation. The agreement to comparative assays is 77.1% (111/144) (95%CI: 69.6-83.2%).

### 6. Precision

The precision of the abia CMV IgM was determined by 20 days × 3 samples × 2 replicates covering the measuring range.

#### Intra-assay (within run) precision

Data	Serum sample #1	Serum sample #2	Serum sample #3
Mean (U/ml)	12.5	3.8	1.9
S <sub>r</sub>	0.4	0.1	0.1
CV (%)	3.5	3.5	5.0

#### Inter-assay (between-run) precision

Data	Serum sample #1	Serum sample #2	Serum sample #3
Mean (U/ml)	12.5	3.8	1.9
S <sub>rr</sub>	0.22	0.11	0.05
CV (%)	1.8	2.9	2.7

## XIII. LIMITS OF THE TEST

- It is inadmissible to make a diagnosis only on the basis of the anti-CMV IgM testing results. The diagnosis of acute CMV infection is possible only in the presence of clinical manifestations and a complex of laboratory studies (detection of an increase in the level of anti-CMV IgG, the detection of high levels of anti-CMV IgM, the isolation of the virus in urine or positive PCR result in serum).

- Poor correlation of results obtained with different commercial kits for IgM testing may be caused by distinctions in immunological responses depending on the type of CMV antigens used.

#### **XIV. CONDITIONS OF STORAGE AND TRANSPORTATION**

Expiry date is indicated on the packaging.

Keep in dark dry place at 2-8 °C. Freezing is prohibited.

Transportation should be done at 2-8 °C. Transportation at 9-25 °C is allowed not more than during ten (10) days.



#### **AB Diagnostic Systems GmbH**

Sportfliegerstraße 4, Berlin, 12487, Germany

Tel. +49 30 208987160, Fax: +49 30 208987199

E-Mail: info@ab-ds.de, www.ab-ds.de

#### **XV. REFERENCES**

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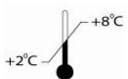





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## XVI. EXPLANATION OF SYMBOLS

	For in vitro diagnostic use		
	Manufacturer		Consult Instruction for use
	Date of manufacture CCYY-MM		Symbol “exclamation mark”
	Catalog number		Symbol “corrosion”
	Sufficient for		Symbol “health hazard”
	Batch code	<b>Danger!</b> <b>Warning!</b>	Signal words
	Storage temperature limitation		Keep away from sunlight
	Expiry date CCYY-MM-DD		Keep dry
	Top		Fragile, handle with care

**Scheme of the assay**

<b>1</b>	<b>Add</b>	90 µl of Preliminary Sample Diluent and 10 µl of the samples (conduct on the plate for preliminary dilution of samples)
<b>2</b>	<b>Add</b>	100 µl of Positive Control, Negative Control
<b>3</b>	<b>Add</b>	90 µl of Sample Diluent
<b>4</b>	<b>Add</b>	10 µl of preliminary diluted samples
<b>5</b>	<b>Incubate</b>	30 min, (37.0 ± 1.0) °C, microplate incubator
<b>6</b>	<b>Wash the plate</b>	Working Washing Solution, not less than 380 µl, 4 times
<b>7</b>	<b>Add</b>	100 µl of Conjugate
<b>8</b>	<b>Incubate</b>	30 min, (37.0 ± 1.0) °C, microplate incubator
<b>9</b>	<b>Wash the plate</b>	Working Washing Solution, not less than 380 µl, 4 times
<b>10</b>	<b>Add</b>	100 µl of TMB-Substrate
<b>11</b>	<b>Incubate</b>	20 min, 18-24 °C in a dark place
<b>12</b>	<b>Add</b>	150 µl of Stopping Reagent
<b>13</b>	<b>Read the optical density</b>	450 nm/620-680 nm